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Fiona O'Mahony

Royal College of Surgeons in Ireland

Warren Thomas

Royal College of Surgeons in Ireland

Brian J. Harvey

Royal College of Surgeons in Ireland

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Novel Female Sex-Dependent Actions of Oestrogen in the Intestine

Fiona O'Mahony, Warren Thomas and Brian J. Harvey

Abstract

The intestine is an oestrogen responsive organ and circulatory oestrogens suppress Cl^- secretion across the epithelium of the colon to promote fluid retention at the estrus stage of the menstrual cycle. Ion transporters in the colon which are involved in Cl^- secretion show differential expression between males and females as do the signalling protein kinase intermediates involved in acutely regulating these transporters. Work from our laboratory has identified the KCNQ1/KCNE3 channel as one of the principal targets for oestrogen-induced signalling cascades in the distal colon. Through inhibition of the KCNQ1 channel, basolateral K^+ recycling is decreased so reducing the favourable electrochemical gradient for Cl^- extrusion at the apical membrane. The actions of oestrogen on non-reproductive tissues such as the colon, kidney, lung and sweat gland will affect whole body electrolyte and fluid homeostasis and also has consequences for reproductive potential.

Rapid and Genomic Ion transporter Responses to E2

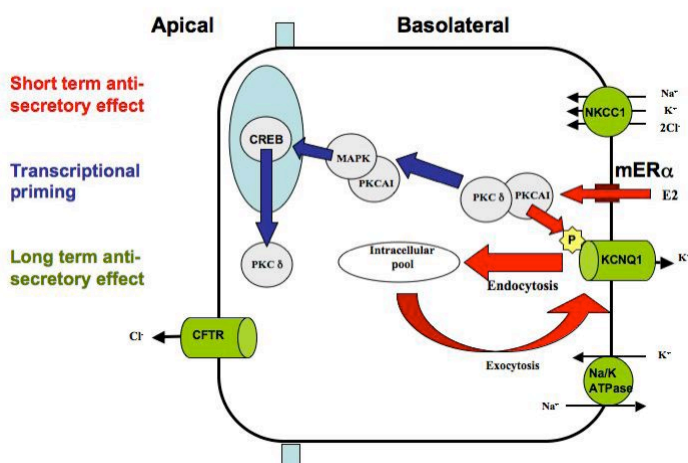


Figure 1. Mechanism of rapid oestrogen inhibition of cAMP activated chloride ion secretion and subsequent induction of fluid retention in the isolated distal colon from female rats. Cl^- ions enter the cell via the basolateral NKCC1 protein and exit via apical CFTR . Na^+ and K^+ recycling occur via the Na^+ K^+ ATPase pump and the KCNQ1 channel at the basolateral membrane. Oestrogen rapidly activates $\text{PKC}\delta$ (2 min), which in turn

activates PKA . The kinase complex translocates to the plasma membrane and associates with the cytoplasmic region of the KCNQ1 channel. PKA induces a rapid serine phosphorylation of the channel. Phosphorylation of the KCNQ1 channel results in an inhibition of K^+ recycling and in turn inhibition of Cl^- secretion. In addition to inducing the rapid and anti-secretory effect, oestrogen induces a transcriptional response via CREB resulting in the up-regulation of expression of $\text{PKC}\delta$ and down-regulation of the expression of ion transporters. In this way the rapid 'non-genomic' effects of oestrogen promote transcriptional priming of signalling intermediates of the anti-secretory response and also feed-forward to sustain the anti-secretion through inhibition of ion transporter expression and activation of inhibitory protein kinases.

Introduction

Oestrogen is the primary female reproductive hormone and exerts biological effects through the modulation of transcriptional and cell signalling processes in target tissues (1). Oestrogen regulates cell growth and differentiation in primary and secondary reproductive organs but also acts on what are generally regarded as non-reproductive tissues expressing oestrogen receptors (ERs). The role of oestrogen in modulating the physiology of non-reproductive tissues is an emerging area of investigation. ERs are expressed in the respiratory tract, kidney, sweat gland and colon as well as at sites in the brain (2). The prominence of secretory and absorptive epithelia among the list of oestrogen target tissues suggests an important role for oestrogen in the regulation of whole body electrolyte and fluid balance through the modulation of ion transport processes (3, 4). Over the course of the oestrous cycle large fluctuations in circulatory oestrogen concentrations are experienced by the female body and these changes modulate the structure and function of reproductive tissues, particularly the uterus to facilitate embryo implantation. Through oestrogen synthesis and release by the ovaries, the ion transport activities of epithelial tissues such as the distal colon are also synchronized with the female reproductive cycle. The importance of oestrogen action on epithelial ion transport to the physiological processes of conception, implantation and pregnancy are still unknown.

The two isoforms of the oestrogen receptor, ER α and ER β , show marked differential tissue expression and biological roles. The receptors act as ligand-dependent transcription factors to positively or negatively regulate the expression of a broad repertoire of oestrogen responsive genes. While bound to oestrogen, these receptors interact with specific DNA sequences to facilitate the assembly of the transcription pre-initiation complexes at gene promoter sites. It is also now clear that ERs interact with adjacent transcription factors at *cis* elements in promoter sequences to regulate transcription in a far more subtle manner than was previously imagined. Reports over the last two decades have shown that oestrogen and other steroid hormones also stimulate the rapid activation of protein kinase signalling cascades in target tissues that potentiate and modulate the direct transcriptional effects of the nuclear ERs. These rapid 'non'genomic' signalling events are initiated at membrane localized oestrogen receptors (5-7) which may be splice variants of the nuclear receptor (8, 9) or may be truncated forms of the ER α associated with the cell membrane through acylation (10). Even though the rapid activation of protein kinases by oestrogen has been established, the biological role of these responses is only now being elucidated. We have identified a role for oestrogen-induced protein kinase signalling cascades in the suppression of fluid secretion in the distal colon which represents a novel and important facet in the hormone's range of physiological effects (11).

Oestrogen and Colonic Physiology

The distal colon is the principal site for water conservation in the body. The transport of ions, particularly Na⁺ absorption and Cl⁻ secretion across the epithelium, determines the rate of water loss and recovery from the lumen of the colon. Disruption of this equilibrium results in conditions such as secretory diarrhoea. The absorption of water from the faeces is coupled to Na⁺ absorption through the Na⁺/H⁺ exchanger or through the epithelial Na⁺ channel (ENaC). Cl⁻ driven secretion serves to hydrate the mucosa and also lubricates the passage of the faecal matter through the colon (12). Cl⁻ is

transported into the epithelial cells of the colon at the basolateral surface through the $\text{Na}^+/\text{K}^+/\text{2Cl}^-$ (NKCC1) co-transporter and exits the cell across the apical membrane into the lumen through the cystic fibrosis trans-membrane conductance regulator (CFTR). The Na^+/K^+ -ATPase at the basolateral membrane provides the driving force for apical Cl^- secretion and ENaC-dependent Na^+ absorption, while the electrochemical gradient is maintained by K^+ recycling through basolateral K^+ channels such as KCNQ1 (13). KCNQ1 plays a crucial role in regulating the rate of salt and water transport across a number of epithelia including that of the small intestine, nephron and colon (14). The transporters engaged in electrolyte transport processes across the colon and other epithelia are subject to precise transcriptional control while acutely acting secretagogues such as forskolin or carbachol modulate transporter activity through the stimulation of signalling cascades coupled to intracellular cAMP or Ca^{2+} , respectively. We have found that oestrogen suppresses the secretagogue-induced Cl^- secretion in the distal colon suggesting that the hormone may also have an anti-secretory role in normal colon physiology (Figure 1) (15). The observation that antagonism of basolateral K^+ recycling also attenuates Cl^- secretion and so mimics the effects of oestrogen in the colon gave an insight into a potential mechanism by which oestrogen could produce its anti-secretory effect (16). Furthermore, in cardiac tissue oestrogen suppresses the KCNQ1-mediated K^+ current suggesting this channel subunit could be a potential molecular target for oestrogen anti-secretory action (17).

Oestrogen and the Colonic Epithelium

The epithelium of the colon expresses both $\text{ER}\alpha$ and $\text{ER}\beta$ (18). However, in the crypts of the proximal colon $\text{ER}\alpha$ is expressed more highly at the base of the crypt while $\text{ER}\beta$ expression is predominant in the mid-section of the crypt and in the lumen surface cells (19). This segregation of ER isoform expression suggests antagonistic roles for the two receptors and differential effects of circulatory oestrogen on the physiological function of the cells located at different sites along the crypt. Progenitor cell proliferation at the base of the crypt gives way to enterocyte differentiation in the midsection and shedding of senescent or apoptotic cells at the lumen surface. $\text{ER}\beta$ has been proposed as a tumour suppressor in colorectal cancer (CRC). $\text{ER}\beta$ expression is selectively lost during tumour progression through methylation-dependent gene silencing (20) and hormone replacement therapy (HRT) reduces the incidence of CRC in postmenopausal women (21, 22). Recent data suggests that HRT may also be protective in suppressing episodes of inflammatory bowel disease in post-menopausal women through the anti-inflammatory effects of oestrogen (23). The role of oestrogen in regulating normal colon physiology and ion transport is less well characterized. However, studies with $\text{ER}\beta$ $-/-$ mice demonstrated a role for the receptor in cell differentiation and the organization of colon tissue architecture through regulation of cell to cell interactions such as tight junction and desmosome assembly (24). In our studies, we have used isolated rat colonic crypts as an experimental model to study the role of oestrogen in suppressing agonist-induced Cl^- secretion and K^+ recycling. KCNQ1 is a voltage-gated, low conductance K^+ channel activated during cAMP-stimulated Cl^- secretion in the colon (25). We found that the activity of this channel was suppressed following oestrogen treatment at physiological levels of the hormone. The oestrogen effect was non-additive to the effects of Chromanol 293B, a specific blocker of KCNQ1, so confirming this channel as an important effector of the oestrogen anti-secretory response. The anti-secretory action of oestrogen is superimposed upon an ion

transporter protein expression profile that differs between males and females. NKCC1, CFTR and Na⁺/K⁺-ATPase α -subunit are all expressed at lower abundance in colonic crypts isolated from female rats compared to males (11). The KCNQ1 channel subunit is not differentially expressed in the male and female colonic epithelium, however the expression of the KCNE3 regulatory subunit is less abundant in females compared to males (11).

Oestrogen-Induced Signalling in the Colon

Oestrogen suppresses forskolin-induced secretion in the distal colon within 10 minutes of hormone treatment suggesting that the effect is at least in part modulated by a signalling cascade rather than by a direct transcriptional response (15). The involvement of a protein kinase signalling cascade was confirmed by the sensitivity of the oestrogen effect to PKC antagonism (15, 16, 26). The phosphorylation of membrane transporters is an important regulatory mechanism contributing either to the direct activation of the transporter or to its localization at the cell membrane. KCNQ1 exists at the cell membrane as part of a multi-molecular complex with the A-kinase anchoring protein Yotiao and a regulatory β subunit. KCNQ1 is phosphorylated at residue Ser27 by membrane associated PKA to modulate its activity in the heart (27). In the distal colon KCNQ1 is in complex with the KCNE3 regulatory subunit. In common with the anti-secretory effect of oestrogen on the colon, the suppression of KCNQ1 activity by oestrogen in the colon was female gender-specific (28). We also found that oestrogen stimulated the rapid activation of PKA in isolated female colonic crypts but not in crypts isolated from male rats (28). The level of PKA phosphorylation induced by oestrogen correlates with the degree of stimulation or suppression of KCNQ1 activity. The activation of different PKA isoforms, which localize to different cellular sites, have different physiological consequences. PKA activation in oestrogen treated colonic crypts is localized to the cytosol rather than to the cell membrane (29). PKA isoform II is anchored to the cell membrane while PKA I is distributed to pools in the cytosol (30). Oestrogen treatment stimulates the physical association of the cytosolic PKA-CI isoform with KCNQ1 whereas the membrane associated PKA is unaffected, The physical interaction of PKA-CI : KCNQ1 correlates with the oestrogen-induced serine phosphorylation of the KCNQ1 channel protein (28). The oestrogen-induced inhibition of Cl⁻ secretion and KCNQ1 activity, and the stimulation of PKA-dependent phosphorylation of KCNQ1 were all antagonized by prior inhibition of PKC δ (28). Recent data points to the differential expression of key protein kinase signalling intermediates in tissues from males as compared to females. At least some of these differentially regulated molecules contribute to steroid-induced responses or are themselves transcriptionally regulated by oestrogen (31). Gender differences in PKC δ abundance has been found in aortic smooth muscle (32) and in the liver (33). The abundance of PKC δ in isolated colonic crypts from rats at the estrus stage of the reproductive cycle was on average four-fold higher in females compared to males, while females at the proestrus stage of the cycle express PKC δ at low levels equivalent to males (28). Through the modulation of protein kinase expression, varying circulating levels of oestrogen determine the sensitivity of particular cell signalling cascades to the rapid actions of the hormone. Oestrogen-induced PKC δ expression effectively creates a positive feed-back loop that amplifies PKC δ -dependent oestrogen rapid and genomic responses at the peak of circulatory levels of oestrogen .

Conclusion

Oestrogen modulates the rate of fluid secretion across the female colonic epithelium over the course of the oestrous cycle. Cl^- and fluid secretion are attenuated at the estrus phase of the cycle by the suppression of basolateral K^+ recycling through the KCNQ1 channel. Oestrogen acutely suppresses KCNQ1 activity through phosphorylation of the channel α subunit, which is dependent on the activation of PKC δ and cytosolic PKA. The expression of PKC δ by the colonic epithelium is modulated throughout the oestrous cycle to amplify the kinase-dependent regulation of KCNQ1 during the estrus phase. Precise regulation of Cl^- secretion by the colon at key stages of the reproductive cycle facilitates fluid conservation at a time that is coincident with uterine restructuring and represents a novel facet to the role of oestrogen in the physiology of reproduction.

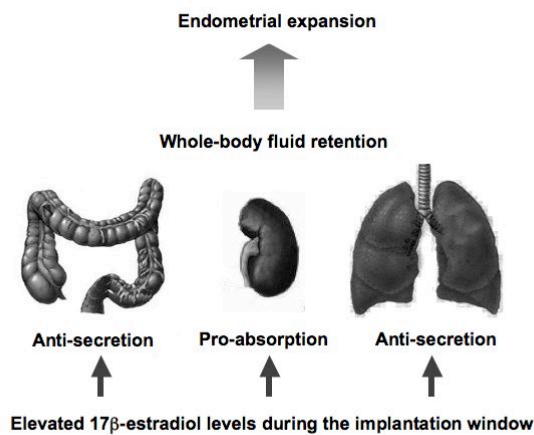


Figure 2. High circulating levels of oestrogen occurring during the implantation window of the estrous cycle (and with hormone replacement therapy and the contraceptive pill) produce an inhibition of chloride ion secretion in the intestine and lung and increase the absorption of sodium in the kidney, resulting in increased fluid retention which is necessary for endometrial expansion and implantation of the blastocyst.

The intestinal tract has previously not been thought of as a sex-steroid targeted organ. Inter-organ interactions have previously been reported between the reproductive, urinary and intestinal systems (22). The identification of functional ER in the intestine indicate a physiological role of oestrogen in the intestine (18). This review highlights the important role for oestrogen in the distal colon in contributing to the regulation of whole body fluid and electrolyte regulation in the female. In preparation for the implantation of the blastocyst, the endometrial lining must expand its vasculature and this is associated with a significant increase in tissue wet weight. High levels of oestrogen result in fluid retention as a result of conservation of large volumes of water from organs such as the kidney, lung and large intestine (Figure 2). This process is a rapid and reversible event as whole body fluid retention of water is cyclical and occurs over short periods of time. In this manner the intestine responds to fluctuating levels of oestrogen to rapidly and dynamically reduce water loss via inhibition of the Cl^- secretory pathway.

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