

1-1-2015

# Developmental control of CFTR: from bioinformatics to novel therapeutic approaches.

Catherine M. Greene

*Royal College of Surgeons in Ireland, [cmgreene@rcsi.ie](mailto:cmgreene@rcsi.ie)*

Dominik Hartl

*University of Tübingen*

---

## Citation

Greene CM, Hartl D. Developmental control of CFTR: from bioinformatics to novel therapeutic approaches. *European Respiratory Journal*. 2015;45(1):18-20

This Article is brought to you for free and open access by the Department of Medicine at [e-publications@RCSI](mailto:e-publications@RCSI). It has been accepted for inclusion in Medicine Articles by an authorized administrator of [e-publications@RCSI](mailto:e-publications@RCSI). For more information, please contact [epubs@rcsi.ie](mailto:epubs@rcsi.ie).

---

— Use Licence —



This work is licensed under a [Creative Commons Attribution-Noncommercial-Share Alike 4.0 License](https://creativecommons.org/licenses/by-nc-sa/4.0/).

---

## **Developmental control of *CFTR*: from bioinformatics to novel therapeutic approaches**

Catherine M. Greene<sup>1</sup> and Dominik Hartl<sup>2</sup>

<sup>1</sup>Dept. Medicine, Royal College of Surgeons in Ireland, Dublin, Ireland.

<sup>2</sup>CF Research Group, Children's Hospital, Dept. I, University of Tuebingen, Tuebingen, Germany.

### Correspondence:

Catherine Greene, Dept. Medicine, RCSI Education & Research Centre, Beaumont Hospital, Beaumont, Dublin 9, Ireland

T: 01-8093712 F: 01-8093808 E: [cmgreene@rcsi.ie](mailto:cmgreene@rcsi.ie)

Celebrating the 25<sup>th</sup> anniversary of identifying the genetic cause of cystic fibrosis, tremendous progress has been made in the understanding and the treatment of this complex, but still fatal disease [1]. Despite advances, several key questions on the origins and pathomechanisms of CF remain poorly understood. For instance, the precise genetic and epigenetic mechanisms that regulate expression and stability of the *cystic fibrosis transmembrane conductance regulator (CFTR)* gene remain incompletely defined.

*CFTR* gene expression is a carefully controlled process that is spatially and temporally regulated. Notwithstanding the extensive research that has been carried out on *CFTR* over the past few decades, developmental control of *CFTR* expression is still poorly understood [2]. Transcription of *CFTR* starts at distinct sites depending on the tissue or developmental stage and is positively regulated by a selection of transcription factors, including C/EBP proteins and FOXA factors. Beyond that, *CFTR* is controlled post-transcriptionally by microRNAs (miRNAs). miRNAs are regulatory factors involved in most biological processes and it is

becoming increasingly clear that they play a key role in the development and manifestations of CF lung disease [3-13]. These small non-coding RNAs act post-transcriptionally to inhibit protein production. Their involvement in the pathogenesis of CF lung disease stems from the fact that their expression is altered *in vivo* in the CF lung due to intrinsic and extrinsic factors; to date defective chloride ion conductance, endoplasmic reticulum stress, inflammation and infection have been implicated in altering endogenous miRNA expression in this setting.

With their recent study, Viart *et al.* now expand substantially our understanding of how *CFTR* expression is regulated [14] (Figure 1). The authors identified regulatory elements that participate in *CFTR* downregulation, particularly FOXA1, FOXA2 and C/EBP $\alpha$ . Using bioinformatics, they further identified four new AU-rich elements (ARE) and experimentally tested their roles in regulation of *CFTR* expression. Following a series of intricate studies, ARE-5698 was found to play a role in destabilizing *CFTR* mRNA. Regarding miRNAs predicted to bind to the *CFTR* 3'UTR, miR-101 had the strongest repressive effect. By comparing miRNA expression profiles of adult and foetal lungs, three specific miRNAs (miR-145, miR-150 and miR-451) were found to have a temporal effect, being significantly up regulated in the adult lung and therefore contributing to downregulation of *CFTR*.

With a view to therapeutics that might enhance *CFTR* expression, miRNA binding-blocker oligonucleotides (MBBOs, also known as target site blockers/protectors or miRNA masks) were designed. The function of an MBBO is to prevent binding of a miRNA to one specific mRNA target. Here MBBOs for several miRNAs that bind the *CFTR* 3'UTR were tested *in vitro* and *ex vivo* using non-CF and p.Phe508del homozygous CF primary human nasal epithelial cells grown at an air-liquid interface. MBBOs targeting the miR-145 and miR-101 sites were most effective, as they stabilised *CFTR* mRNA and enhanced *CFTR* protein expression. Finally, both MBBOs significantly enhanced *CFTR* function in CFBE41o- cells.

This report adds significantly to our existing knowledge regarding miRNA regulation of *CFTR* [5-10, 13] and the developing field of miR-based *CFTR* therapeutics [15]. The data confirms previous studies implicating miR-145 and miR-101 as important modulators of *CFTR* expression [5, 6, 8, 9] and build on these by demonstrating how MBBOs based on these miRNAs can affect *CFTR* gene expression and *CFTR* protein function. The authors suggest that the MBBOs may be developed as tools for *CFTR* correction in people with CF. Indeed this is an appealing option given that MBBOs are less likely to have off-target effects compared to some other miRNA inhibition strategies [reviewed in 16].

Although there are a series of novel findings from this paper, the work also raises a number of important questions for further investigation. The CF lung is inherently associated with microbial colonisation. Whether infection impacts upon the newly described transcriptional and post-transcriptional mechanisms controlling *CFTR* expression remains elusive. This will be particularly important for the further development of the MBBOs as therapeutics for CF. Previous studies have reported how infection and inflammation affect the expression of miRNAs, including those that regulate *CFTR* [9, 13]. This suggests that higher than normal levels of MBBOs may be required to ensure an inhibitory effect on endogenous miRNA activity. Nonetheless it will be a very exciting development when miRNA-modulating drugs for CF advance to the stage of clinical studies. A second point worth considering is the relevance of Viart *et al.*'s findings beyond CF. Separate sets of studies have clearly shown that cigarette smoke affects *CFTR* and miRNA expression. Although the mechanisms responsible for altered *CFTR* expression in, for example, the COPD lung are not restricted to miRNA-mediated effects [17], miR-101 in particular is known to be increased by cigarette smoke extracts (CSE) and directly impacts on *CFTR* expression [8]. Finally, it is becoming increasingly evident that dysfunctional *CFTR* as a result of cigarette smoking can contribute to the pathophysiology of a range of extrapulmonary disease conditions, such as

chronic pancreatitis, male infertility and cachexia [18]. Expanding upon the findings from airway epithelial cells reported here into non-lung cells that express CFTR may yield novel therapies for these disease conditions beyond CF.

## FIGURE LEGENDS

**Figure 1.** *Transcriptional and post-transcriptional regulation of CFTR.* In the adult lung (A) the transcription factors FOXA1/2 and C/EBP $\alpha$  negatively regulate *CFTR* gene expression whilst (B) ARE-5698 destabilises *CFTR* mRNA and miR-145 and miR-101 block *CFTR* expression. MBBOs targeting the miR-145 and miR-101 sites can restore *CFTR* expression.

## References

- [1] Mall MA, Hartl D. CFTR: cystic fibrosis and beyond. *Eur Respir J*. 2014 Jun 12. pii: erj02280-2013. [Epub ahead of print]
- [2] McCarthy VA, Harris A. The CFTR gene and regulation of its expression. *Pediatr Pulmonol*. 2005; 40: 1-8.
- [3] Oglesby IK, Bray IM, Chotirmall SH, Stallings RL, O'Neill SJ, McElvaney NG, Greene CM. miR-126 is downregulated in cystic fibrosis airway epithelial cells and regulates TOM1 expression. *J Immunol*. 2010; 184:1702-1709.
- [4] Bhattacharyya S, Balakathiresan NS, Dalgard C, Gutti U, Armistead D, Jozwik C, Srivastava M, Pollard HB, Biswas R. Elevated miR-155 promotes inflammation in cystic fibrosis by driving hyperexpression of interleukin-8. *J Biol Chem*. 2011; 286: 11604-11615.
- [5] Gillen AE, Gosalia N, Leir SH, Harris A. MicroRNA regulation of expression of the cystic fibrosis transmembrane conductance regulator gene. *Biochem J*. 2011; 438: 25-32.
- [6] Megiorni F, Cialfi S, Dominici C, Quattrucci S, Pizzuti A. Synergistic post-transcriptional regulation of the Cystic Fibrosis Transmembrane conductance Regulator (CFTR) by miR-101 and miR-494 specific binding. *PLoS One* 2011; 6: e26601.
- [7] Ramachandran S, Karp PH, Jiang P, Ostedgaard LS, Walz AE, Fisher JT, Keshavjee S, Lennox KA, Jacobi AM, Rose SD, Behlke MA, Welsh MJ, Xing Y, McCray PB Jr. A microRNA network regulates expression and biosynthesis of wild-type and DeltaF508 mutant

cystic fibrosis transmembrane conductance regulator. *Proc Natl Acad Sci U S A.* 2012; 109: 13362-13367.

[8] Hassan F, Nuovo GJ, Crawford M, Boyaka PN, Kirkby S, Nana-Sinkam SP, Cormet-Boyaka E. MiR-101 and miR-144 regulate the expression of the CFTR chloride channel in the lung. *PLoS One.* 2012; 7: e50837.

[9] Oglesby IK, Chotirmall SH, McElvaney NG, Greene CM. Regulation of cystic fibrosis transmembrane conductance regulator by microRNA-145, -223, and -494 is altered in  $\Delta F508$  cystic fibrosis airway epithelium. *J Immunol.* 2013; 190: 3354-3362.

[10] Amato F, Seia M, Giordano S, Elce A, Zarrilli F, Castaldo G, Tomaiuolo R. Gene mutation in microRNA target sites of CFTR gene: a novel pathogenetic mechanism in cystic fibrosis? *PLoS One.* 2013; 8: e60448.

[11] Megiorni F, Cialfi S, Cimino G, De Biase RV, Dominici C, Quattrucci S, Pizzuti A. Elevated levels of miR-145 correlate with SMAD3 down-regulation in cystic fibrosis patients. *J Cyst Fibros.* 2013; 12: 797-802.

[12] Rupani H, Sanchez-Elsner T, Howarth P. MicroRNAs and respiratory diseases. *Eur Respir J.* 2013; 41: 695-705.

[13] Ramachandran S, Karp PH, Osterhaus SR, Jiang P, Wohlford-Lenane C, Lennox KA, Jacobi AM, Praekx K, Rose SD, Behlke MA, Xing Y, Welsh MJ, McCray PB Jr. Post-



transcriptional regulation of cystic fibrosis transmembrane conductance regulator expression and function by microRNAs. *Am J Respir Cell Mol Biol.* 2013; 49: 544-551.

[14] Viart V, Bergougnoux A, Bonini J, Varihl J, Chiron R, Tabary O, Molinari N, Claustres M, Taulan M. TFs and miRNAs that regulate fetal to adult CFTR expression change are new targets for CF. *Eur Resp J* In Press

[15] Amato F, Tomaiuolo R, Nici F, Borbone N, Elce A, Catalanotti B, D'Errico S, Morgillo CM, De Rosa G, Mayol L, Piccialli G, Oliviero G, Castaldo G. Exploitation of a very small peptide nucleic acid as a new inhibitor of miR-509-3p involved in the regulation of cystic fibrosis disease-gene expression. *Biomed Res Int.* 2014; 2014: 610718.

[16] Hassan T, McKiernan PJ, McElvaney NG, Cryan SA, Greene CM. Therapeutic modulation of miRNA for the treatment of proinflammatory lung diseases. *Expert Rev Anti Infect Ther.* 2012; 10: 359-368.

[17] Rasmussen JE, Sheridan JT, Polk W, Davies CM, Tarran R. Cigarette smoke-induced Ca<sup>2+</sup> release leads to cystic fibrosis transmembrane conductance regulator (CFTR) dysfunction. *J Biol Chem.* 2014; 289: 7671-7681.

[18] Raju SV, Jackson PL, Courville CA, McNicholas CM, Sloane PA, Sabbatini G, Tidwell S, Tang LP, Liu B, Fortenberry JA, Jones CW, Boydston JA, Clancy JP, Bowen LE, Accurso FJ, Blalock JE, Dransfield MT, Rowe SM. Cigarette smoke induces systemic defects in cystic fibrosis transmembrane conductance regulator function. *Am J Respir Crit Care Med.* 2013; 188: 1321-1330.

